Multiple resistance to glyphosate and imazethapyr in *Bidens subalternans*

Resistência múltipla a glyphosate e imazethapyr em *Bidens subalternans*

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**ABSTRACT**

Glyphosate has been widely used to control greater beggarticks populations that are resistant to acetolactate synthase (ALS) inhibitors in South America. However, herbicide control failures has been observed over the last three growing seasons in grain production areas of Paraguay. In this research, we report the first case of multiple resistance to glyphosate (EPSPS) and imazethapyr (ALS) in greater beggarticks (*Bidens subalternans*) population from Paraguay. This conclusion was supported by dose-response experiments conducted in two consecutive years (2018 and 2019) with a putative resistant (R) and a susceptible (S) population. Alternative herbicides were also tested for post-emergence control of R population. For glyphosate, the resistant factor (RF) values were 8.8- (2018) and 15.7-fold (2019). For imazethapyr, the RF values were 59- and 58-fold, in 2018 and 2019, respectively. Treatments with 2,4-D, dicamba, 2,4-D + glyphosate, dicamba + glyphosate, lactofen, fomesafen, ammonium-glufosinate, atrazine, and bentazon provided more than 80% control of the R population. This is the first case of multiple resistance to glyphosate and imazethapyr in greater beggarticks (*Bidens subalternans*) in the world. The mechanisms underlying resistance in this biotype should be evaluated in future research.

**Index terms:** Greater beggarticks; EPSPS inhibitor; ALS inhibitors; chemical control.

**INTRODUCTION**

Glyphosate targets the shikimate pathway, causing plant death through 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) enzyme inhibition (Duke; Powles, 2008). Glyphosate is a non-selective, systemic and low-cost herbicide widely used in different crop production systems (Heap; Duke, 2018). The frequency in glyphosate use significantly increased after the commercialization of glyphosate-resistant (GR) crops, leading to the evolution of 43 unique GR cases (Heap, 2019). In South America, 17 species have been identified as GR, including 9 mono- and 8 dicotyledonous species (Heap, 2019).

Acetolactate synthase (ALS) inhibiting herbicides are normally sprayed in low rates, have broad spectrum, low toxicity for humans, and are selective to many crops (Yu; Powles, 2014). These herbicides are toxic to plants by inhibiting valine, leucine, and isoleucine amino acids production. Weeds normally evolve resistance to ALS inhibitors more rapidly than to other herbicides probably
due to the high frequency of target site mutations affecting herbicide binding to the enzyme also due to herbicide metabolization by P450 monoxygenases enzymes (Prentson; Powles, 2002). Likewise, the number of weeds that are resistant to ALS inhibitors is greater than any other herbicide group (Heap, 2014).

Species belonging to the genus *Bidens* L. (Asteraceae), usually known as beggarticks, are widespread across South American areas in tropical and subtropical environments, infesting roadsides and agricultural crops (Grombone-Guaratini; Solferini; Semir, 2004). In grain fields, greater beggarticks interference with density of more than 40 plants m⁻¹ can cause up to 34% in yield losses (Galon et al., 2016). While the genus *Bidens* is composed by many species, only two of them are considered important weeds: *Bidens subalternans* and *B. pilosa* (Grombone-Guaratini; Solferini; Semir, 2004). These two species share high levels of morphological similarities, but they can be differentiated by the number of seed awns, 2 to 3 in *B. pilosa* and 3 to 4 in *B. subalternans*, and the type of leaf branches in the third top of the plants, dichotomous in *B. pilosa* and alternates in *B. subalternans* (Kissmann; Groth, 2000).

*Bidens* species have a long herbicide resistance history in South America. The first case was reported in *B. pilosa* resistant to ALS inhibitors infesting soybean fields in the beginning of 1990’s (Christoffoleti, 2002). Then, *B. subalternans* was also identified as resistant to ALS inhibitors in the other soybean crop in 1996. (Lamego et al., 2009). Multiple resistance to ALS inhibitors and atrazine has also been reported for both species in double-crop soybean/corn fields in Brazil (Takano et al., 2016; Heap, 2019). More recently, GR *B. pilosa* was reported in México (Cruz et al., 2016).

Over the last three growing seasons, growers have reported greater beggarticks (*Bidens subalternans*) survival after glyphosate treatment in soybean and corn areas in Paraguay. Considering that extensive soybean and corn fields areas are infested with greater beggarticks resistant to ALS inhibitors (Mendes et al., 2019), it is possible that some populations have been selected for multiple resistance (glyphosate and ALS inhibitors) in these areas. Therefore, our objective was to investigate multiple resistance to EPSPs- and ALS- inhibitors in a putative resistant greater beggarticks population from Paraguay, and evaluate alternative herbicides to chemical control for that population.

**MATERIAL AND METHODS**

Greater beggarticks (*B. subalternans*) seeds were collected in January 2017, in a soybean field at Santa Rosa del Monday (25°56’45’’ S 54°55’22’’ W), Paraguay. The species was confirmed as *B. subalternans* due to morphological characteristics. The sampled field had previous history of soybean in the fall/summer followed by corn in the autumn/winter and the area had been sprayed with glyphosate for at least 10 years in soybean and five years in corn. Seeds were sampled from 20 random plants that survived glyphosate application. Subsequently, seeds were cleaned, stored in paper bags, identified and kept in room temperature until the experiments were conducted.

Seeds were sowed in 200-cell flats (1 x 1 x 3 cm) filled with potting soil (MacPlant®). After germination, seedlings were transplanted to 1 dm³ pots filled with soil (27% clay, pH 6.2 and organic matter 2.1%) and potting soil at 1:1 ratio. Thirty plants (three-leaf stage, 10-12 cm height) were sprayed with glyphosate at 960 g ae ha⁻¹ (Roundup Transorb R®, 2 L ha⁻¹) and imazethapyr at 106 g ai ha⁻¹ (Zethapyr® 1,0 L ha⁻¹) in a tank mix. After application of both herbicides, surviving plants were kept in pots to produce seeds. Flats and pots were kept under greenhouse conditions with daily irrigation of 5 mm. Seeds from treated plants (F1) were collected and stored as previously described.

Dose-response experiments were conducted with the F1 generation of putative resistant (R) and another population previously determined as susceptible (S) collected at Maringá (PR). Plant growth conditions were conducted as describe above. One experiment was designed for each herbicide in a 2x8 factorial (two populations and eight herbicide doses). Glyphosate was applied at 0, 120, 240, 480, 960, 1920 and 3840 g ae ha⁻¹ on S and at 0, 240, 480, 960, 1920, 3840 and 7680 g ae ha⁻¹ on R population. For imazethapyr, S and R were treated with 0, 13.2, 26.5, 53, 106, 212, 424 and 848 g ai ha⁻¹. The label dose for *B. pilosa* used as field dose for *B. subalternans* control is 480-720 g ae ha⁻¹ for glyphosate and 106 g ai ha⁻¹ for imazethapyr. The treatments were applied on plants at the three-leaf stage (10-12 cm), and the experimental design was completely randomized with four replications. Dose-response experiments were conducted in 2018 and repeated in 2019.

An additional experiment was conducted to evaluate the efficacy of alternative herbicides on the R population. Plants were grown in the greenhouse as described above. Different herbicides that are currently recommended for greater beggarticks control in soybeans and corn post emergence were evaluated (Rodrigues; Almeida, 2018). The treatments were: untreated check; 2,4-D at 670 g ae ha⁻¹ (DMA®, 1 L ha⁻¹); dicamba at 480 g ae ha⁻¹ (Atecreta®, 1 L ha⁻¹); 2,4-D + glyphosate at 608 +
960 g ae ha⁻¹; dicamba + glyphosate at 480 + 960 g ae ha⁻¹; lactofen at 180 g ai ha⁻¹ (Naja®, 0.75 L ha⁻¹); fomesafen 250 g ai ha⁻¹ (Flex®, 1 L ha⁻¹); ammonium-glufosinate at 400 ai g ha⁻¹ (Finale®, 2 L ha⁻¹); chlorimuron-ethyl at 20 g ai ha⁻¹ (Classic®, 80 g ha⁻¹), atrazine at 1,500 g ai ha⁻¹ (Proof®, 3 L ha⁻¹); mesotrione at 144 g ai ha⁻¹ (Callisto®, 0.3 L ha⁻¹); tembotrione at 76 g ai ha⁻¹ (Soberan®, 0.18 L ha⁻¹) and bentazon at 720 g ai ha⁻¹ (Basagran®, 1.2 L ha⁻¹). Adjuvants were added following the label of each herbicide.

All applications were performed with a CO₂ pressurized backpack sprayer and a 1.5 m long bar with three ST 100.02 spray nozzles (0.5 m between nozzles, Teejet Technologies®). The sprayer was operated at 195 kpa, providing a spray volume of 150 L ha⁻¹. For all experiments, plant survival was evaluated following a 0-100% scale, in which zero means plant death and 100% means no visual symptoms. At the end of each experiment, shoots were collected and dried at 60 °C for four days and dry mass was measured. Evaluations were performed at 21 days after treatment (DAT) for dose-response experiments and at 14 and 28 DAT for the alternative chemical control experiment.

Dose-response data from each year (2018 and 2019) were subjected to ANOVA (p<0.05) and then to a combined analysis with multiple years for each herbicide. Non-linear regression models with four (glyphosate – Equation 1) or three parameters (imazethapyr – Equation 2) were fit using SigmaPlot 12 software.

\[
\hat{y} = d + \frac{a-d}{1 + \left(\frac{x}{c}\right)^b} \quad (1)
\]

\[
\hat{y} = \frac{a}{1 + \left(\frac{x}{c}\right)^b} \quad (2)
\]

Where \(a\) is the upper limit, \(b\) is the slope around \(c\), \(c\) is the dose to 50% of plant survival or dry mass relative to untreated check (relative dry mass) and \(d\) is the lower limit. The \(x\) values were independent (dose) and \(y\) values were dependent (survival or dry mass) variables. After models adjustment, the dose for 50% of survival (LD₅₀) and the dose for 50% of relative dry mass (ED₅₀) were calculated through inverse equation, by using \(x\) as a function of \(y\) (Ritz et al., 2015). Resistant factors (RF) were obtained using LD₅₀ R/S or ED₅₀ R/S ratio. For the alternative control experiment, data were subjected to ANOVA (p<0.05) and Tukey test (p<0.05) were used to compare means. Both analyses were ran using SISVAR 12 software (Ferreira, 2011).

**RESULTS AND DISCUSSION**

There was a significant interaction (p<0.05) between the two years of experiments for glyphosate and imazethapyr in all variables, therefore, the dose-response curves were fit separately for each year (2018 and 2019).

For glyphosate, LD₅₀ value was higher in R than in S population. The R population was 8.8-fold (2018) and 15.7-fold (2019) more resistant than S, based on the RF values (Figure 1). To achieve efficient control (≤10% survival) on the R population, it would be necessary 4,850 g ae ha⁻¹ based on the data from 2018. On the other hand, even the highest dose tested (7,680 g ae ha⁻¹) was not enough to provide less than 21% survival in 2019. Labeled rates of glyphosate for greater beggarticks range from 480 to 960 g ae ha⁻¹ (Rodrigues; Almeida, 2018), which emphasizes the resistance to glyphosate in the R population (Table 1). Dry mass (ED₅₀) values provided RF of 10- (2018) and 22-fold (2019) (Figure 1 and Table 1), which is in agreement with the survival data.

For hairy beggarticks (B. pilosa), a closely related species, glyphosate resistance levels based on ED₅₀ (20.4-fold) were very similar to ours (Cruz et al., 2016). High RF levels in glyphosate resistance (>10) has also been found in other dicotyledonous weeds, such as Conyza sumatrensis (Santos et al., 2014) and Amaranthus palmeri (Gonçalves Netto et al., 2016).

The R population demonstrated significant resistance levels to imazethapyr (Figure 2). The R population was 59.4-fold (2018) and 58.7-fold (2019) more resistant than S based on survival. Based on the ED₅₀, these values were 24.2-fold in 2018 and 32.4-fold in 2019 (Table 2). In both years, survival levels were ≥62% for R, and the calculated dose of imazethapyr to provide ≤10% survival exceeded the highest dose applied (>848 g ai ha⁻¹). The results found here are similar to the RF range found for ALS inhibitors in other B. subalternans populations from South America (Lamego et al., 2009; Mendes et al., 2019).

Prior to the GR crops era, ALS inhibitors were often used for weed control in pre- and post-emergence, contributing to the selection pressure and evolution of resistant populations around the world (Green, 2014). After the introduction of GR crops, the selection pressure imposed by glyphosate applications increased substantially.
on plants that were already resistant to ALS inhibitors, leading to the evolution of multiple resistant weeds (Heap and Duke, 2018). Similar to greater beggarticks, other weeds have already been identified with multiple resistance to glyphosate and ALS inhibitors, such as *A. palmeri* (Gonçalves Netto et al., 2016), *Lolium rigidum* (Owen et al., 2007), *C. sumatrensis* (Santos et al., 2014) and *Kochia scoparia* (Beckie et al., 2013).

![Figure 1](image)

**Figure 1:** Dose-response curves for glyphosate in greater beggarticks (*Bidens subalternans*). Circles represent the resistant (R) population and squares represent the susceptible (S) population. Bars represent standard errors. Model fit: Plant survival or relative dry mass = \( d + \frac{a-d}{1+(dose/c)^b} \).

**Table 1:** Model parameters, dose in g ae ha\(^{-1}\) to provide 50% of plant survival (LD\(_{50}\)), dose to provide 50% of relative dry mass (ED\(_{50}\)), and resistant factor (RF) in greater beggarticks (*Bidens subalternans*) resistant (R) and susceptible (S) to glyphosate.

<table>
<thead>
<tr>
<th>Variable</th>
<th>Population</th>
<th>a</th>
<th>b</th>
<th>c</th>
<th>d</th>
<th>R(^2)</th>
<th>LD(<em>{50}) or ED(</em>{50}) (^{1/2})</th>
<th>RF(^{2/3})</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>2018</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Survival</td>
<td>R</td>
<td>169.1</td>
<td>0.5</td>
<td>3,856.5</td>
<td>-69.7</td>
<td>0.98</td>
<td>650</td>
<td>8.8</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>100.2</td>
<td>3.8</td>
<td>74</td>
<td>-0.2</td>
<td>0.99</td>
<td>73.8</td>
<td></td>
</tr>
<tr>
<td>Relative dry mass</td>
<td>R</td>
<td>113.9</td>
<td>0.6</td>
<td>466.8</td>
<td>-14.1</td>
<td>0.98</td>
<td>312</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>100.7</td>
<td>1.7</td>
<td>34.7</td>
<td>-0.7</td>
<td>0.99</td>
<td>31.2</td>
<td></td>
</tr>
<tr>
<td><strong>2019</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Survival</td>
<td>R</td>
<td>79.4</td>
<td>0.9</td>
<td>868.4</td>
<td>21.6</td>
<td>0.94</td>
<td>1665.1</td>
<td>15.7</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>102.7</td>
<td>1.4</td>
<td>112.2</td>
<td>-3.4</td>
<td>0.98</td>
<td>106.1</td>
<td></td>
</tr>
<tr>
<td>Relative dry mass</td>
<td>R</td>
<td>135.0</td>
<td>0.4</td>
<td>2,906.8</td>
<td>-37.7</td>
<td>0.66</td>
<td>610</td>
<td>22.1</td>
</tr>
<tr>
<td></td>
<td>S</td>
<td>102.1</td>
<td>1.0</td>
<td>28.8</td>
<td>-2.18</td>
<td>0.99</td>
<td>27.6</td>
<td></td>
</tr>
</tbody>
</table>

Model fitted parameters: Plant survival or relative dry mass = \( d + \frac{a-d}{1+(dose/c)^b} \); \(^{1/2}\) calculated by inverse equation; \(^{2/3}\) LD\(_{50}\) R/S or ED\(_{50}\) R/S ratio.
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Variable Population \(a\) \(b\) \(c\) \(R^2\) \(LD_{50}\) or \(ED_{50}\) \(RF_{2/\_}\)
---
Survival R 94.1 1.6 841.2 0.96 778.9 59.4
S 99.5 0.7 13.3 0.96 13.1
Relative dry mass R 100.4 0.2 231.8 0.93 236 24.2
S 99.8 0.8 9.8 0.98 9.7

Model fitted parameters: Survival or relative dry mass = \(a/1+(dose/c)^b\); calculated by inverse equation; \(2LD_{50}\) R/S or \(ED_{50}\) R/S ratio.

Resistance to glyphosate in weeds has been associated with both target-site (TSR) and non-target-site (NTSR) mechanisms. Target site mutations by single nucleotide polymorphisms are known to confer resistance by changing the enzyme structure, which prevents glyphosate from binding to EPSPS (Sammons;
Another TSR mechanism is the capacity of resistant biotypes to produce more copies of the EPSPS gene compared to susceptible plants, making glyphosate rates insufficient to inhibit the increased enzyme number (Gaines et al., 2010; Wiersma et al., 2015). NTSR mechanisms are often found in weeds and they are normally associated with reduced uptake and translocation, enhanced metabolism, vacuolar sequestration and/or rapid tissue death (Sammons; Gaines, 2014; Moretti et al., 2018). Usually, single mutations in positions Thr-102 or Pro-106 of EPSPS do not result in high RF values (Han et al., 2016). On the other hand, the simultaneous presence of Thr-102-Ile and Pro-106-Leu in resistant plants (TIPS mutation) confer much higher RF values (more than 100-fold) (Yu et al., 2015). The increase in EPSPS copy number and NTS mechanisms also can lead to high RF to glyphosate in weeds (Gaines et al., 2010). In the hairy beggarticks population resistant to glyphosate from México, Cruz et al. (2016) found a Pro-106-Leu in one population and the TIPS mutation in another population, providing RF of 2.4 and 20.7, respectively.

Recently, the mechanism of resistance in a GR B. subalternans from Paraguay was under investigation and the double mutation TIPT (Thr-102-Ile and Pro-106-Thr) was found in one EPSPs allele of two EPSPs homologous in this species (Takano et al., 2019).

Resistance to ALS inhibitors has been widely studied for a long time. Mutations in ALS is the most common mechanism and can occur in eight different amino acids positions of the gene (Heap, 2019). Typically, mutations that result in the highest RF for imidazolinones, such as imazethapyr, are described in Ala-122, Ala-205 and Trp-574 (Powles; Yu, 2014; Heap, 2019). The Trp-574-Leu substitution has been found in a greater beggarticks population from Brazil, resulting in 166-fold resistance to imazethapyr (Lamego et al., 2009). More rarely, enhanced metabolism by P450 monoxygenases enzymes or reduced translocation have also been described in the literature (Guo et al., 2015; Nakka et al., 2017). Investigating herbicide resistance mechanisms to ALS inhibitors in weeds are critical to develop management strategies.

When herbicides were applied on plats at the three-leaf stage (10-12 cm height), most treatments provided good performance on the R population, except chlorimuron-ethyl, mesotrione and tembotrione (Table 3). We have not conducted a dose response for other ALS inhibitors herbicides, but these results suggest that the R population could be cross-resistant to other herbicides.

### Table 3: Efficacy of alternative herbicides to control glyphosate- and imazethapyr-resistant greater beggarticks (Bidens subalternans) in post-emergence.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (g ha⁻¹)</th>
<th>Survival (%)</th>
<th>Relative dry mass (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Untreated check</td>
<td>-</td>
<td>100 a</td>
<td>100 a</td>
</tr>
<tr>
<td>2,4-D</td>
<td>670</td>
<td>20 d</td>
<td>0.7 e</td>
</tr>
<tr>
<td>Dicamba</td>
<td>480</td>
<td>24 d</td>
<td>0.7 e</td>
</tr>
<tr>
<td>2,4-D + glyphosate</td>
<td>670 + 960</td>
<td>20 d</td>
<td>1 e</td>
</tr>
<tr>
<td>Dicamba + glyphosate</td>
<td>480 + 960</td>
<td>20 d</td>
<td>0.7 e</td>
</tr>
<tr>
<td>Lactofen</td>
<td>180</td>
<td>0.2 e</td>
<td>0 e</td>
</tr>
<tr>
<td>Fomesafen</td>
<td>250</td>
<td>0.2 e</td>
<td>0 e</td>
</tr>
<tr>
<td>Ammonium-glufosinate</td>
<td>400</td>
<td>2.2 e</td>
<td>0 e</td>
</tr>
<tr>
<td>Chlorimuron-ethyl</td>
<td>20</td>
<td>77 b</td>
<td>56 c</td>
</tr>
<tr>
<td>Atrazine</td>
<td>1,500</td>
<td>1.2 e</td>
<td>0 e</td>
</tr>
<tr>
<td>Mesotrione</td>
<td>144</td>
<td>51 c</td>
<td>34 d</td>
</tr>
<tr>
<td>Tembotrione</td>
<td>76</td>
<td>59 c</td>
<td>88 b</td>
</tr>
<tr>
<td>Bentazon</td>
<td>720</td>
<td>0.7 e</td>
<td>0 e</td>
</tr>
</tbody>
</table>

Means followed by the same letters indicates no difference by Tukey’s test (P < 0.05); dose in g ha⁻¹; applied with non-ionic surfactant at 0.2% v v⁻¹; applied with mineral oil at 0.5% v v⁻¹; DAT: days after treatment.
in this class, consistent with what has been described in other greater beggarticks populations across South America (Lamego et al., 2009; Mendes et al., 2019). For HPPD-inhibiting herbicides (mesotrione and tembotrione), frequently used in corn fields, their tank mix with atrazine can provide efficient greater beggarticks control (Matte et al., 2018). Despite the levels of control observed with PSII inhibitors (atrazine and bentazon), atrazine resistance has already been reported in this species (Heap, 2019), than, careful recommendations should be made for the sustainable use of PSII herbicides.

Treatments containing 2,4-D, dicamba, and ammonium-glufosinate are important options for burndown weed control and will probably be even more important with the launch of new herbicide tolerance traits in South America, such as Enlist®, Xtend® and Liberty Link®. The results provide evidence that at least four different MoA are still effective on the R greater beggarticks population (synthetic auxins, protoporphyrinogen-oxidase (PPO)-inhibitors, PSII- inhibitors and glutamine-synthetase (GS)-inhibitor) (Table 3). Other options in pre-emergence that were not evaluated in this research may also provide tools to improve greater beggarticks management (e.g. sulfentrazone and flumioxazin) (Lopez Ovejero et al., 2013). Herbicide rotation with different MoAs is essential to prevent and manage multiple resistance evolution. More importantly, additional non-chemicals practices should also be included in weed management strategies, such as crop rotation, cover crops, mechanical and cultural control (Marochi et al., 2018). Thus, machinery cleaning is another important measure to prevent the spread of this problem (Takano et al., 2018). Finally, survey programs should be implemented to monitor the eventual spread of glyphosate R populations of greater beggarticks towards other countries.

CONCLUSIONS

This research confirmed the first case of glyphosate resistance in greater beggarticks (Bidens subalternans) worldwide. This population is also resistant to imazethapyr. Alternative herbicides from at least four different MoA, applied in post-emergence, demonstrated efficient control of this multiple resistant greater beggarticks population.

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